

0040-4020(93)EOO36-F

FUNCTIONALIZED CHLOROENAMINES IN AMINOCYCLOPROPANE SYNTHESIS - XIII.' AZAANNULATED CYCLOPROPANES - RIGID BUILDING BLOCKS FOR OLIGOAMINES

Jens Seibel, Elmar Vilsmaier', Karin Frohlich, Gerhard Maas and Rolf Wagemann

Fachbereich Chemie der Universitat Kaiserslautern, Erwin-Schroedinger-Strase, D-67663 Kaiserslautern, Germany

Abstract: Oligoamines 5, 6 and 7 with rigid 3-azabicyclo[3.l.Olhexyl building blocks were synthesized from di(chloroenamines) 8 and nucleophiles. Sodium borohydride as nucleophile led to endo,endo-tetramines 5a,b; the same stereochemical result generating 5d and 5f was observed for cyanide or methyllithium as reagents. Methylmagnesium bromide reacted with 8 to give mainly exo,exo-tetramine 7f besides small amounts of isomers 5f and 6f. Basicity, conformation and molecular flexibility of the new tetramines 5 - 7 were studied. X-Ray structural analyses pointed out a meander shape of tetramine 5f and a linear arrangement of tetramine 7f.

Putrescine (I), spermidine (2) and spermine (3), naturally occurring oligoamines, strongly influence cell proliferation processes. 2.3 Interactions of these oligoamines with DNA, therefore, were explored intensively.^{4,5} Structural properties of the oligoamines are important in the **formation of the ammonium DNA - phosphate complexes. Oligoamines possessing rigid building blocks should be of interest in this context. Our investigations about rigid diamines with an** azabicyclohexane skeleton^{1,6-9} prompted us to synthesize analogous oligoamines 4 and to **study their properties. Two azabicyclo[3.l.O]hexane units are connected by a piperazine moiety in compounds of type 4 for which three diastereomers 5, 6 and 7 must be considered.**

Interest was applied specially to endo,endo-diastereomers 5 since here molecular flexibility should be mostly restricted. Difunctional chloroenamines 8 were provided as starting materials. Reaction of 8 with sodium borohydride or cyanide should leads,7 to endo,endo-oligoamines 5 $(R¹ = H, CN)$; methylmetal compounds and di(chloroenamine) 8 were chosen as starting materials for a practicable access⁸ to diastereomers 5, 6 and 7 (R¹ = CH₃).

SYNTHESIS OF 1,4-DI-(3-AZABICYCLO[3.1 .OlHEXYL)-PlPERAZlNES 5, 8 AND 7

Diienamines) 1la.b were easily obtained from piperidinones 9a,b and piperazine (IO) by a standard procedure. Chlorination of 1 la,b with N-chlorosuccinimide (12) in dichloromethane at

8a, 9a. Iia: R2 = Me 8b. gb, lib: R2 = Rzl

-50°C gave **di(chloroenamines) 8 (8a: 49%; 8b:** 70% **yield). The monochlorination of both** enamine units of 11 generating 8 was clearly established by the ¹³C NMR data of the products $indicating only one single $C = C$ -double bond [8a: 143.0 (s), 103.1 (d); 8b: 143.2 (s), 102.8]$ **(d)) and only one single CHCI-moiety [8a: 53.3 (d); 8b: 53.5 (d)l.**

Reaction of di(chloroenamines) 8a,b with sodium borohydride in acetonitrile provided bicyclic compounds: Thereby, di(borane-adduct) 13a was isolated in 43% yield as product of starting material 8a. Excessive interaction of 1 N aqueous hydrochloric acid (80°C, 24 h) was necessary for deboronation of 13a to give 5a (75% yield). The corresponding N-benzyl borane adduct 13b was hydrolyzed very easily; here, acidic destroying of excess borohydride cleaved already the borane - amine complex leading to oligoamine 5b (48% yield) as isolable reaction product. N-unsubstituted cyclopropanopyrrolidine 5c (85% yield) could be obtained by hydrogenolysis of N-benzyl oligoamine 5b in methanol in the presence of Pd/C.

Reaction of di(chloroenamine) 8a with sodium cyanide provided bicyclic dinitrile 5d (84% yield) which could be reduced to hexamine 5e by lithium aluminum hydride (64% yield).

Di(chloroenamine) 8a was transformed into pure endo,endo-tetramine 5f (33% yield) by methyllithium (14); diastereomers 8f or 7f could not be detected 1H NMR spectroscopically in the crude reaction mixture. The analogous interaction of methylmagnesium bromide (15) with di(chloroenamine) 8a led to a mixture of three diastereomers 5f, 6f and 7f. The latter was **produced with remarkable selectivity (50% yield of pure isomer 7f); isomers 5f (5% yield) and 6f (11% yield) were isolated as pure compounds by chromatography.**

Formation of endo,endo-tetramine 5f as only isolable product from the chloroenamine methyllithium reaction was in accordance with the exclusive generation of diamines 17 from chloroenamine 16 and methyllithium (14). Methylmagnesium bromide (15), however, showed **different stereoselection in the reaction with chloroenamine 16 on the one hand (gave equal** amounts of 17 and 18)⁸ and di(chloroenamine) 8a on the other hand.

CONFIGURATION OF 1,4-DI-(3-AZABICYCLO[3.1 .OlHEXYL)-PIPERAZINES 5, 6 AND 7

The number of the ¹³C NMR signals indicated the highly symmetric structure of the bicyclic **compounds 5, 7 and 13a: Only two signals at all were found for the cyclopropane moieties and only one triplet appeared for the piperazine methylene groups of 5, 7 and 13a. Different configurations of the two azabicyclohexyl moieties in compound 6f caused two sets of signals for the constitutionally identical piperazine substituents.**

endo-Piperazine configuration of 5b,c and 13a was established by the ³J_{HH} coupling of the triplet of the C(6)-H-signal (5b: $3J_{HH} = 6.0$ Hz; 5c: $3J_{HH} = 6.8$ Hz; 13a: $3J_{HH} = 6.1$ Hz). **These values are characteristic of coupling of syn H-atoms of a cyclopropane ring system (e.g. ref.7 and references cited therein). The configuration of 5a [C(G)-H-signal superposed by a multiplet] was deduced on the basis of the configuration of the corresponding precursor 13a.** endo-Piperazine configuration of 5d / 5e was indicated by the magnitude of the ³J_{CH} coupling of 4.1 Hz splitting the nitrile ¹³C NMR signal of 5d into a triplet (e.g. ref.⁶ and references cited **therein).**

X-Ray structural analyses showed the endo,endo-configuration of 5f (Fig. 1) and the exo,exoconfiguration of 7f (Fig. 2) and allowed indirectly the assignment of the endo,exo-configuration of the third isomer 6f. The ¹³C NMR δ -values of the C-methyl moiety in 5f and 7f are in accordance with this structural information: Highfield shifting of this signal $(\delta = 3.9 \text{ ppm})$ **corresponds to compound 7f with an endo-methyl moiety and lowfield shifting of this signal (6 = 14.7 ppm) is characteristic of isomer 5f with the methyl group in exo-position.**

The plots of the X-ray structural analyses indicate a meander type arrangement (isomer 5f) or a linear shape (isomer 7f) of the new type of oligoamines depending on the configuration of the 3-azabicyclo[3.1 .O]hexyl building blocks.

CONFORMATION AND BASICITY OF 1,4-DI-(3-AZABlCYCLO[3.1 .O]HEXYL)-PIPERAZINES 5, 6 AND 7

X-Ray analyses demonstrate clearly the presence of a chair conformation for endo,endo-isomer 5f and a boat conformation for exo,exo-tetramine 7f in the solid state. This result for isomer 5f is in accordance with an X-ray structural analysis⁹ of 17; the boat conformation, found for 7f, is confirmed by a ¹H NMR spectroscopic conformational study with diamine⁹ 18. The interplanar angles C(1)C(2)C(4)C(5) / C(2)N(2)C(4) of 5f (24.0^o) and 7f (28.6^o) show a clear **buckling of the pyrrolidine system in both cases.**

Fig. 1 Schakal-plot¹⁰ of 1,4-Bis-(1*a,*5*a,6β*-3,6-dimethyl-3-azabicyclo[3,1.0]hex-6-yi **plperazine (5f)**

Fig. 2 Schakal-plot ¹⁰ of 1,4-Bis-(1*a*,5*a*,6*a*-3,6-dimethyl-3-azabicyclo[3.1.0]hex-6-yi **piperazine (7f)**

Table 1 Selected bond distances, N,N-distances,^a torsional angles and interplanar angles^a of 1,4-bis-(3,6-dimethyl-3-azabicyclo[3.1.0]hex-6-yl)-piperazine diastereomers 5f and 7f

a The numbering of the nitrogen atoms in Fig. 1, Fig. 2 and Table 1 in this paper was partially changed with respect to the numbering in the deposited data for better comparison with other systems.- b H(2)_A / H(4)_{A'} are in the endo-position and H(2)_B / H(4)_B' are in the exo-position of **the 3-azabicyclo[3.1 .O]hexane system.**

A potential influence of the conformation of the pyrrolidino ring on the geometry of the cyclopropane system of a 3-azabicyclo[3.1.O]hexyl compound can be excluded by comparing the bond lengths C(1)-C(5), C(1)-C(6) and C(5)-C(6) of 5f and 7f.

The conformation of the 3-azabicyclo[3.l.O]hexane units in the oligoamines 5, 6 and 7 in solution could be deduced from ¹H NMR data: A "zero-coupling" between H_A/H_A·(endo-Hatoms) and H_x/H_x. indicates⁹ the presence of a boat conformation (clearly found for 5c, 7f and one azabicyclohexyl moiety of 6f); highfield shifting of H_A/H_A·(endo-H-atoms) and a visible

coupling between H_A/H_{A} (endo-H-atoms) and H_X/H_{X} , on the other hand, are characteristic⁹ of **a chair conformation (clearly found for 5a, 5b, 5d, 58, 5f and one azabicyclohexyl moiety of 6f).** The dihedral angles $H(1)$ -C(1)-C(2)-H(2)_A / $H(4)$ _A-C(4)-C(5)-H(5) in 5f (chair) and 7f (boat) **from the X-ray structural analyses (Table I) underline the correctness of the conformational** analysis on the basis of the coupling between $H_A/H_{A'}$ and $H_X/H_{X'}$. In the case of 5c the ¹H **NMR data additionally informed about the axial position of the hydrogen atom at the N(3)** nitrogen atom. The ¹H NMR signal of the pyrrolidine CH₂-unit of 5c gave a further splitting at -28°C (toluene) due to coupling with the N(3)-H-atom. Values of $3J_{HH} = 7.0$ Hz for H_A/H_{A'} and ${}^{3}J_{HH}$ = 11.2 Hz for H_B/H_B, are consistent only with an axial N(3)-H-atom.

Basicity of the tetramines 5, 6 and 7 was studied in water at 25°C, aqueous hydrochloric acid **(1 N for 5a. c, d; 0.1 N for 5f, 6f and 7f) was used for the titration in water. pK,-values were** determined by the application of the Henderson - Hasselbalch equation¹¹ at the corresponding **half neutralization points leading to the simple expression: pH = pK,. The pH of aqueous solutions was measured with a combined glass electrode; aqueous buffer solutions of pH 4.0,** 7.0 and 9.0 were used for the calibration. The used concentrations (c_o) and the pK_a-values are **given in Table 2.**

a 50 mL of the solution were used for each titration.- b Limit of error for pK_a-units: \pm 0.02.- c **Uptake of one proton.- d Uptake of two protons.**

For all tetramines with two identical azabicyclohexyl moieties (5 and 7) only one single step appeared in the titration curves corresponding to the uptake of two protons. The titration plot of 6f, however, showed a very slight bending in the area of the first neutralization point and a sharp step for the uptake of the second proton. Hexamine 6e gave two clear steps in the titration curve corresponding to two protons each. The uptake of only two protons by tetramines 5 - 7 and the basicity differences of 5f, 6f and 7f correspond quite well to the behaviour of diamines¹ 17 and 18. Twofold protonation of 5 - 7 and the uptake of the first two **protons in 5e should take place at the pyrrolidine nitrogen atoms N(3) and N(3'1.12 Deactivation of the piperazine moiety in 5** - **7 by the cyclopropane ring (see ref. 1) prevents futher protonation in the aqueous system. It is known, that introduction of a cyano group into the** *a***position of an aminoalkane decreases basicity by about 5 pK-units (e.g. a-aminoacetonitrile's** pK_a: 5.3; methanamine¹³ pK_a: 10.7). A cyano group interacts as electron deficient moiety also with an adjacent cyclopropane system¹⁴; this generates a situation in 5d which is similar to a **vinylogous a-aminonitrile and which makes understandable the decrease of basicity of 5d with respect to that of 5a or 5f.**

Tertiary amines generally are less strong bases than the secondary analogues. In contrast to this, N-methyl compound 5a showed the same basicity as the corresponding N-H-derivatives 5c. This unexpected behaviour can be attributed to conformational differences of 5a' (chair conformation) and 5c (boat conformation).

MOLECULAR FLEXIBILITY OF 1,4-DI-(3-AZABICYCLO[3.1 .O]HEXYL)-PIPEFIAZINES 5, 6 AND 7

Free activation enthalpy ΔG^+ of topomerization of the hydrogen atoms of the piperazine unit **gave an insight into the molecular flexibility of the oligoamines 5, 6 and 7 possessing rigid** azabicyclohexyl building blocks. The ΔG^* -values for this topomerization process were obtained **by temperature dependent 1H NMR spectroscopy. In the case of tetramines 5, a change of the** piperazine H-signals from an AA'XX'-system into an A₄-system should be observed. The 400 MHz spectra allowed an easy determination of δH_A and δH_X due to a symmetrical shape of each of the signal pattern. The ΔG ⁺-values were estimated by application of the formula for **the coalescence of uncoupled signals. 1s The applicability of this approximation in the case of a piperazine system was demonstrated with N,N'-dimethylpiperazine: A value of 55.7 kJ/mol** (H_A/H_A) : 2.49 ppm; H_X/H_X : 2.09 ppm; T_c = 285 K; $C_A D_B C D_A$), thus obtained, agrees sufficiently well with the value of 55.6 kJ/mol (CD₂Cl₂) reported by Petrakis.¹⁶ A change from **an ABXY system to an AA'XX' system for the piperazine 1H NMR signals could be observed for tetramine 6f with increasing temperature. Due to partial superposition of the signals, the AG* values also were determined with the approximation formula for the exchange of noncoupling protons.'5**

Compound	T [K]	H _{AA'} or H_A , H_Y [ppm]	$H_{XX'}$ or H_B , H_X [ppm]	T_c [K] [kJ/mol]	∆G ⁺ ª
5а	230	2.45	2.22	265	52.9
5c	245	2.29	1.77	295	57.1
5d	270	2.46	2.06	330	64.9
Бе	290	2.83	2.37	340	66.6
5f	270	2.58	2.27	332	66.0
6f	282	2.47 ^b	2.27c	313	63.2
		2.62 ^d	2.49e	310	63.7
7f	220	2.61	2.52	242	50.0

Table 3. AG * - Values of the dynamics of the piperazine ring of compounds 5a, 5c, 5d, 5f, 6f and 7f determined on the basis of 1H NMR data (400 MHz) and coalescence temperatures (T,) in CsDsCDs.

a Calculated with the approximation formula for the uncoupled case (ref.¹⁵).- b H_Y.- c H_X.- d HA.- e Hs.-

The rigid 3-azabicyclo[3.1.0]hexyl moieties at the piperazine unit of 5, 6 and 7 act as **equatorial anchoring groups leading to definite geometries in all cases. Rotation of the azabicyclo(3.1 .Olhexyl units at the piperazine group additionally can be hindered in the case of their fixation in endo position in compounds 5. As expected, hindrance of this rotation depends** on the substituent R¹ [no additional hindrance for $R^1 = H$ (see ref.⁷) and moderate additional hindrance for $R^1 = CN$ (see ref.⁶) and $R^1 = Me$ (see ref.⁸)]. Thus, meander type tetramines 5 with further substituents in 6,6'-position (R¹ = H) should be the most interesting representants **of the new oligoamines.**

EXPERIMENTAL

tH NMR and 1aC NMR spectra were obtained with a Bruker AMX 400 spectrometer (TMS as internal standard). IR spectra were measured on a Perkin-Elmer 397 Infrared Spectrophotometer. A MAT 90 (Finnigan) spectrometer was used for mass spectra. Microanalyses were performed with a Perkin-Elmer 2400 Elemental Analyzer. The amines were titrated with a Metrohm Titrino SM 702 apparatus using Metrohm electrodes (combined pHglass electrode with Ag/AgCI/KCI (3 N) as inner reference electrode].

Piperidone Enamine 11 - General Procedure: A solution of piperidinone 9 (0.1 mol, 9a: 11.3 g; 9b: 18.9 g), 4-toluenesulfonic acid (0.2 g, 1.05 mmol) and piperazine (10) (4.3 g, 0.05 mol) in **benzene (150 mL) was heated in a Dean-Stark apparatus for 11 h. Concentration of the solution to 20 mL and standing, at room temperature gave crystalline enamines 11 which were isolated by suction and washed with ice-cold ether.**

1,4-Bis-(1,2,3,6-tetrahydro-l-methyl-pyridin-4-yl~-piperazine **(lla): Yield: 12.1 g (88%), mp 118°C; IR (KBr, cm⁻¹) 1640 (C=C); ¹H NMR (CDCI₃)** δ **2.25 (4H), 2.55 (4H) (AA'BB'-system), 2.34 (s, 6H), 2.85 (s, 8H), 2.98 (m_c, 4H), 4.62 (t, 2H); ¹³C NMR (CDCI₃) δ 143.2 (s), 97.8 (d),** 54.1 (t), 52.2 (t), 47.3 (t), 45.5 (q), 27.7 (t). Anal. Calcd for C₁₆H₂₈N₄: C, 69.52; H, 10.21; **N, 20.27. Found: C, 69.5; H, 10.4; N, 20.5.**

1,4-Bis-(1-benzyl-1,2,3,6-tetrahydro-pyridin-4-yl)-piperazine (11b): Yield: 20.1 g (93%), mp **128°C; IR (KBr, cm⁻¹) 1640 (C=C); ¹H NMR (CDCI₃)** δ **2.21 (4H), 2.58 (4H) (AA'BB'-system), 2.85 (s, 8H), 3.06 (m,, 4H), 3.57 (s, 4H), 4.60 (t, 2H), 7.247.36 (m, IOH); 13C NMR (CDCI3) d 143.6 (s), 138.4 (s), 129.1 (d), 128.1 (d), 126.9 (d), 97.7 (d), 62.7 (t), 52.7 (t), 49.8 (t),** 47.3 (t), 27.8 (t). Anal. Calcd for C₂₈H₃₆N₄: C, 78.46; H, 8.47; N, 13.07. Found: C, 78.4; H, *8.4; N, 13.1.*

Di(chloroenamines) 8 - General Procedure: A solution of N-chlorosuccinimide (12) (5.34 g, 40 **mmol) in dichloromethane (140 mL) was dropped at -50°C during 1 h to a stirred solution of enamine (20 mmol, lla: 5.53 g; 1 lb: 8.57 g) in dichloromethane (60 mL). Stirring was** continued at -50^oC for 1 h. Then cooling was removed to warm up the mixture to room **temperature. Succinimide was removed by extraction with saturated aqueous sodium carbonate solution (3 x 80 mL). Removing the solvent in vacua and trituration of the residue with pentane (150 mL) gave pure chloroenamines 8. Recrystallization from ether led to colorless crystals.**

1,4-Bis-(3-chloro-1,2,3,6-tetrahydro-l-methyl-pyridin-4-yl/-piperazine **(8a): Yield: 3.38 g (49%). mp 148°C (decomp.); IR (KBr, cm⁻¹) 1640 (C=C); ¹H NMR (CDCI₃)** δ **2.82 (H_{B1}, 2H), 2.95** (H_{A1}, 2H), 4.80 (H_{X1}, 2H) (ABX-system), 2.85 (H_{B2}, 2H), 3.30 (H_{A2}, 2H), 4.61 (H_{X2}, 2H) (ABX-system), 2.39 (s, 6H), 2.83-2.91 (m, 4H), 3.00-3.09 (m, 4H); ¹³C NMR (CDCl₃) *δ* 143.0 (s), 103.1 (d), 60.8 (t), 54.3 (t), 53.3 (d), 47.5 (t), 45.2 (q). Anal. Calcd for $C_{16}H_{26}Cl_2N_4$: C, **55.65; H, 7.59; N, 16.23. Found: C, 55.8; H, 7.7; N, 16.3.**

1,4-Bis-(l-benzyl-3-chloro- 1,2,3,6-tetrahydro-pyridin-4-yl)-piperazine **(8b): Yield: 6.96 g (70%),** mp 155°C (decomp.); IR (KBr, cm⁻¹) 1640 (C=C); ¹H NMR (CDCl₃) δ 2.94-3.08 (H_{A2}, H_{B1}, 4H and m, 4H), 3.33 (H_{A1}, 2H), 4.80 (H_{X1}, 2H), 2.81 (H_{B2}, 2H), 4.59 (H_{X2}, 2H) (2 ABX-systems), **2.85-2.92 (m, 4H), 3.56 (Ha,, 2H). 3.73 (HA,, 2H) (AB-system), 7.23-7.40 (m, IOH); 3C ' NMR (CDC13) ,j 143.2 (s), 137.7 (s), 128.9 (d), 128.2 (d), 127.1 (d), 102.8 (d), 61.6 (t), 57.9 (t),**

53.5 (d), 52.6 (t), 47.4 (t). Anal. Calcd for C₂₈H₃₄Cl₂N₄: C, 67.60; H, 6.89; N, 11.26. Found: **C, 67.2; H, 7.0; N, 10.8.**

Reaction of Di(chloroenamines) 8 with 6odium Borohydride - General Procedure: A suspension of di(chloroenamine) 8 (10 mmol; 8a: 3.45 g; 8b: 4.97 g) and sodium borohydride (3.78 g, 100 mmol) in acetonitrile (120 mL) was stirred at 70°C for 100 h. Excess sodium borohydride was removed by suction; the solvent was evaporated and the residue dissolved in water (50 mL). Aqueous hydrochloric acid (I N) was added at O°C to the solution till pH 1. The acidic solution was stirred till the end of hydrogen evolution. Basification by aqueous sodium hydroxide (1 N) till pH 10 and extraction with ether (3 x 80 mL) gave crude products 13a and 5b, respectively. Pure products were obtained by crystallization from acetonitrile.

[6,6'-(Piperazine- 1,4-diyll/bis-ltrih ydro-l la,5a,6\$-3-meth yl-Sazabic ycJol3. 1. *Ojhexane-Ns,N?lboron* **(13a): Yield: 1.30 g (43%), mp 273°C; IR (KBr, cm⁻¹) 2220-2400 (B-H); ¹H NMR** $(CDCl₃)$ δ 1.92 (H_Y, $3J_{XY} = 3J_{XY} = 6.1$ Hz, 2H), 2.01 (H_X, H_X, 4H), 2.55 (H_A, H_A, 4H), 3.39 (H_B, H_R, 4H) (AA'BB'XX'Y-system), 2.64 (s, 6H), 2.30-2.70 (m, 8H); ¹³C NMR (CDCI₃) δ 61.2 (t), 51.9 (t), 49.7 (q), 49.6 (d, ¹J_{CH} = 160 Hz), 26.1 (d, ¹J_{CH} = 175 Hz). Anal. Calcd for **C1sH34B2N4: C, 63.20; H, 11.27; N, 18.42. Found: C, 63.0; H, 11.2; N, 18.4.**

1,4-Bis-(la,5a,6fl-3-benzyl-3-azabicyc/o13.1.OIhex-6-yll-piperazine **(5b): Yield: 2.05 g (48%),** mp 135°C; ¹H NMR (CDCI₃) δ 1.71 (H_X, H_X, 4H), 1.78 (H_Y, $3J_{XY} = 3J_{XY} = 6.0$ Hz, 2H), 2.29 **(HA, HA,, 4H), 3.14 (Ha, Ha., 4H) (AA'BB'XX'Y-system), 2.50 (s, broad, 8H), 3.66 (s, 4H),** 7.22-7.35 (m, 10H); ¹³C NMR (CDCI₃) *δ* 140.0 (s), 128.7 (d), 128.1 (d), 126.6 (d), 58.8 (t), **52.1 (t), 52.0 (t), 48.2 (d,** 1 **J_{CH} = 167 Hz), 24.7 (d,** 1 **J_{CH} = 171 Hz). Anal. Calcd for C2sH3sN4: C, 78.46; H, 8.47; N, 13.07. Found: C, 78.1; H, 8.7; N, 13.2.**

1,4-Bis-(la,5u,6&3-methyl-3-azabicyclo[3.l .O]hex-6-yl)-piperazine (5a): Di(borane) adduct 13a (1.30 g, 4.3 mmol) was added to a mixture of aqueous hydrochloric acid (1 N, 60 mL) and acetonitrile (10 mL) and stirred at 80°C for 24 h. The solvent was evaporated in vacua, the residue was dissolved in water (30 mL) and brought to pH 12 by addition of aqueous sodium hydroxide (5 N). Extraction with dichloromethane (3 x 20 mL) at O°C gave crude 5a which was recrystallized from acetonitrile. Yield: 0.88 g (75%), mp 140° C; ¹H NMR (CDCI₃) δ 1.70-1.80 (H_X, H_X, H_Y, 6H), 2.24 (H_A, H_A, 4H), 3.26 (H_B, H_B, 4H) (AA'BB'XX'Y-system), 2.32 (s, 6H), **2.45 (s, broad, 8H).** ¹³C NMR (CDCI₃) δ 53.8 (t), 52.0 (t), 48.3 (d, ¹J_{CH} = 165 Hz), 40.3 (q), 25.3 (d, ¹J_{CH} = 170 Hz); MS (70 eV) m/e = 277.0 ([M + 1]⁺, 10%), 190.6 (93%), 94.2 (100%). Anal. Calcd for C₁₆H₂₈N₄: C, 69.52; H, 10.21; N, 20.27. Found: C, 69.7; H, 10.1; N, **20.5.**

1.4-Bis-(1a.5a.6B-3-azabicyclo[3.1.0]hex-6-yl)-piperazine (5c): A solution of di(benzyl) compound 5b (0.86 g, 2.0 mmol) in methanol (I 50 mL) was added to Pd/C catalyst (10%) (0.40 g) in a hydrogen atmosphere and stirred for 24 h. The reaction was stopped when the theoretical amount of hydrogen (90 mL) was consumed. Removal of the catalyst by filtration and evaporation of the methanol gave crude 5c which was recrystallized from acetonitrile. Yield: 0.42 g (85%), mp 215°C (decomp.); ¹H NMR (CDCI₃) δ 1.45-1.53 (m, 4H), 1.76 (t, $3J_{HH}$ = 6.8 Hz), 1.90-2.85 (m, unsplit, 10 H), 2.91-3.06 (m, 8H). ¹³C NMR (CDCI₃) δ 53.8 (t), 48.1 (t), 44.8 (d, ¹J_{CH} = 162 Hz), 23.1 (d, ¹J_{CH} = 169 Hz). Anal. Calcd for C₁₄H₂₄N₄: C, **67.70; H, 9.74; N, 22.56. Found: C, 67.4; H, 9.6; N, 22.5.**

6.6'-(Piperazine-1.4-divl)-bis-(1*a.5a.68-3-methyl-3-azabicyclo(3.1.0)hexane-6-carbonitrile)* (5d): **Di(chloroenamine) 8a (3.45 g, 10 mmol) and sodium cyanide (1.23 g, 25 mmol) were heated in a mixture of acetonitrile (80 mL) and water (8 mL) to 50-60°C for 16 h. Then the solvent was** removed in vacuo and the residue was dissolved in dichloromethane (60 mL). Washing the **solution with water (2 x 50 mL) and evaporating the dichloromethane gave crude dinitrile 5d which was recrystallized from acetonitrile. Yield: 2.73 g (84%). mp 231°C (decomp.); IR (KBr,** cm⁻¹) 2110 (C \equiv N); ¹H NMR (CDCI₃) δ 2.27 (H_A, H_A, 4H), 2.33 (H_X, H_X, 4H), 3.32 (H_B, H_B, **4H) (AA'BB'XX'-system), 2.29 (s, 6H), 2.62-2.76 (m, 8H). ¹³C NMR (CDCI₃)** *δ* **117.3 (t, ³J_{CH}** $=$ 4.1 Hz), 53.2 (t), 49.2 (t), 43.7 (s), 40.1 (q), 34.0 (d, ¹J_{CH} = 175 Hz). Anal. Calcd for **C,sH,sNs: C, 66.23; H, 8.03; N, 25.74. Found: C. 66.4; H, 8.3; N, 25.8.**

1,4-Bis-{1a,5a,6*β*-6-(aminomethyl)-3-methyl-3-azabicyclo[3.1.0]hex-6-yl}-piperazine (5e): Lithi**um aluminum hydride (1.14 g, 30 mmol) was added to a suspension of dinitrile 5d (0.49 g, 1.5 mmol) in ether (100 mL). The mixture was refluxed for 3 d. Excess lithium aluminum hydride was destroyed at -20°C by aqueous potassium hydroxide (20%, 30 mL). Insoluble crude hexamine 5e was obtained by filtration at room temperature and washing with water (5 mL)** and ether (5 mL). Extraction with acetonitrile (90 mL) at 80°C, concentration of the filtrate to **60 mL and cooling at O°C gave pure crystals of 5e. Yield: 0.32 g, (64%), mp 198OC; 1H NMR** (CDCI₃) δ 0.94 (broad, unsplit, 4H), 1.75 (m_c, 4H), 2.17 (m_c, 4H); 2.30 (s, 6H), 2.55 (H_X, H_X, 4H), 2.85 (H_A, H_A, 4H) (AA'XX'-system), 2.75 (s, 4H), 3.14 m_c, 4H); ¹³C NMR $(CDCl₃)$ δ 55.2 (s), 54.2 (t), 50.1 (t), 43.5 (t), 40.4 (q), 32.4 (d, ¹J_{CH} = 167 Hz). Anal. Calcd **for C,sH34N6: C, 64.63; H, 10.25; N, 25.12. Found: C, 64.6; H, 10.0; N, 24.8.**

1,4-Bis-(1a,5a,6*β*-3,6-dimethyl-3-azabicyclo[3.1.0]hex-6-yl)-piperazine (5f) from Di(chloroenamine) 8a and Methyllithium (14): An ethereal solution of methyllithium (14) (1.6 M; 18 mL, 28.8 mmol) was dropped at -20°C within 1 h to a mixture of di(chloroenamine) 8a (1.0 g, 2.9 **mmol) in ether (100 mL). Stirring was continued at -2O'C for 10 h, then the suspension was** warmed up till 10°C very slowly (4 h). The crude reaction mixture was hydrolyzed by addition of ice (20 g) and of H₂SO₄ (95%, 3 mL). The clear solution was extracted with ether (3 x 30 **mL), basified by saturated aqueous NaOH solution till pH = 14 was reached and extracted**

with ether (200 mL) in a Kutscher-Steudel apparatus for 4 d. Drying the ethereal solution with MgS04, removal of the ether by evaporation and distillation of the residue in a Kugelrohr apparatus at 100-l 10°C / 0.0001 Torr gave pure tetramine 5f as colorless crystals. Yield: 0.29 g (33%), mp 185°C (decomp.); ¹H NMR (CD₃C₆D₅) δ 0.83 (s, 6H), 1.36 (H_{X1}, H_{X'1}, 4H), 2.17 (H_{A1}, H_{A'1}, 4H), 3.17 (H_B, H_{B'}, 4H) (AA'BB'XX'-system), 2.35 (s, 6H), 2.27 (H_{X2}, H_{X'2}, 4H), 2.58 (H_{A2}, H_{A'2}, 4H) (AA'XX'-system). ¹³C NMR (CDCI₃) δ 54.3 (t), 49.4 (s), 48.5 (t), 40.3 (q), 35.1 (d, ¹J_{CH} = 166 Hz), 14.7 (q). Anal. Calcd for C₁₈H₃₂N₄: C, 71.01; H, 10.59; N, **18.40. Found: C, 70.8; H, 10.5; N, 18.3.**

Reaction of Di(chloroenamine) 8a with Methylmagnesium Bromide (15): An ethereal solution of **methylmagnesium bromide (15) (3 M; 20 mL, 60 mmol) was dropped at room temperature within 1 h to a mixture of di(chloroenamine) 8a (1.0 g, 2.9 mmol) in ether (100 mL). The suspension was refluxed for 4 d. Then the crude reaction mixture was worked up as described above. The ethereal solution from the Kutscher-Steudel extraction was evaporated to give a mixture of the 3 diastereomeric tetramines 5f, 6f and 7f. The crude amines were dissolved in** ether (30 mL); standing for 16 h at 4°C gave crystalline exo, exo-tetramine 7d which was **isolated by suction (0.40 g, 45%). The amines in the remaining solution were separated by** chromatography (column: ϕ : 2 cm, length: 20 cm; basic $A₁O₃$). Elution with ether gave **further exo,exo-isomer 7f as first and endo,exo-isomer 6f as second fraction. endo,endo-Tetramine 5f was obtained by subsequent elution with methanol.**

l,CBis-(la, 5a. **6f3-3,6-dimeth** *yl-3-azabicyclol3. l.Olhex-6- yll-piperazine* **(5f) Yield: 0.043 g** (5%); mp 185^oC (decomp.); 1H NMR data identical with those of 5f which was obtained from **the reaction of 8a with methyllithium (14).**

l-/la,5a,6fi-3,6-Dimeth yl-3-azabicyclol3.1.Olhex-6- yll-4-/la,5a, 6a-3,6-dimethyl-3-azabicyclo-[3.1.O]hex-6-yl)-piperazine **(6f)** Yield: 0.096 g, 11%), mp 112°C; ¹H NMR (CD₃C₆D_E) δ 0.80 (s, 3H), 1.37 (s, 3H), 1.33 (H_{X1}, H_{X1}, 2H), 2.18 (H_{A1}, H_{A1}, 2H), 3.18 (H_{B1}, H_{B1}, 2H) $(AA'BB'XX'$ -system), 1.41 $(H_{X2}, H_{X'2}, 2H)$, 2.89 $(H_{A2}, H_{A'2}, 2H)$, 2.46 $(H_{B2}, H_{B'2}, 2H)$ **(AA'BB'XX'-system), 2.16 (s, 3H), 2.34 (s, 3H), 2.30 (broad, 2H), 2.50 (broad, 4H), 2.60 (broad, 2H). 13C NMR (CDCIs) d 55.5 (t), 53.7 (t), 49.8 (~1, 48.4 (t), 48.2 (t), 47.0 (s), 41.5** (q) , 40.1 (q), 34.7 (d, ¹J_{CH} = 169 Hz), 31.2 (d, ¹J_{CH} = 169 Hz), 14.7 (q), 3.6 (q). Anal. **Calcd for C,sH32N,: C, 71.01; H, 10.59; N, 18.40. Found: C, 71.2; H, 10.8; N, 18.5.**

1,4-Bis-(la,5a,6a-3,6-dimethyl-3-azabicyclo[3.1 .Olhex-6-ylkpiperazine (7f) **Yield: 0.443 g** (50%), mp 195°C; ¹H NMR (CD₃C₆D₅) δ 1.34 (s, 6H), 1.42 (H_{X1}, H_{X'1}, 4H), 2.87 (H_{A1}, H_{A'1}, 4H), 2.44 (H_B, H_{B'}, 4H) (AA'BB'XX'-system), 2.15 (s, 6H), 2.50 (s, 8H). ¹³C NMR (CDCl₃) *δ* 55.0 (t), 48.5 (t), 47.2 (s), 41.4 (q), 31.4 (d, ¹J_{CH} = 168 Hz), 3.9 (q). Anal. Calcd for **C,sHs2N4: C, 71.01; H, 10.59; N, 18.40. Found: C, 71.1; H, 10.4; N, 18.4.**

X-Ray Crystal Structure Analysis of 5f. 17.18 Single crystals of 5f were obtained by **crystallization from ether.**

Crystal data: $C_{18}H_{32}N_A$, F.W. = 304.5; triclinic, space group P; χ a = 6.375(3), b = 6.470(6), **c** = 11.115(4) \hat{A} ; α = 97.34(5), β = 101.29(3), γ = 94.99(5)°; V = 442.9(9) \hat{A}^3 ; 1 molecule per unit cell; $D_x = 1.142$ g \cdot cm⁻³; crystal size 0.60 \times 0.50 \times 0.15 mm.

Data collection: Diffractometer Enraf-Nonius CAD 4, monochromatized Mo-K_n radiation; 1567 independent reflections with $4.00 < 2\theta < 50.00^{\circ}$ [w/20 scan, scan width $(0.95 + 0.35 \tan)$ **O)', scan speed 1.8 - 5.0 o** . **min-ll, no absorption correction.**

Structure solution and refinement: Full matrix least-squares method; H atoms refined isotropically, 997 reflections with $I_{obs} > 2 \sigma(I_{obs})$; 164 variables, unit weights, weighting scheme w = $4 \cdot F_{obs}^2/[\sigma(I)]^2 + (P \cdot F_{obs}^2)^2$, P = 0.015; maximum shift/error ratio 0.03, R = 0.046, $R_w = 0.038$.

X-Ray Crystal Structure Analysis of 7f. 1711s Single crystals of 7f were obtained by crystallization from ether.

Crystal data: $C_{18}H_{32}N_A$, F.W. = 304.5; monoclinic, space group P2₁/c ; a = 9.986(3), b = 5.830(2), c = 15.491(22) \hat{A} ; $\alpha = \gamma = 90$, $\beta = 95.74(5)$ °; V = 897.3(19) \hat{A}^3 ; 2 molecules **per unit cell;** $D_x = 1.127$ g cm⁻³; crystal size 0.70 x 0.50 x 0.30 mm.

Data collection: Diffractometer Enraf-Nonius CAD 4, monochromatized Mo-K_n radiation; 1741 independent reflections with 4.00 < 20 < 50.00° [w/20 scan, scan width (0.85 + 0.35 tan O)^o, scan speed 2.5 - 4.0^o · min⁻¹], no absorption correction.

Structure solution and refinement: Full matrix least-squares method; H atoms refined isotropically, 1164 reflections with $I_{obs} > 2 \sigma (I_{obs})$; 164 variables, unit weights, weighting scheme w = $4 \cdot F_{obs}^2/[\sigma(I)]^2 + (P \cdot F_{obs}^2)^2$, P = 0.015; maximum shift/error ratio 0.09, R = 0.053, $R_w = 0.050$.

Acknowledaments: Support of this work by the Deutsche Forschungsgemeinschaft is greatfully acknowledged. Additionally the work was sponsored by the Fonds der Chemischen Industrie. We want to thank Dr. Heike Slodzyk for mass spectra measurement.

REFERENCES AND NOTES

- **1. Foregoing paper: Butz, V.; Vilsmaier, E.; Maas, G.** *J. Chem. Sot. Perkin Trans. 2,* **1993, in press.**
- *2.* **Morris, D. R.; Marton, L. J. Polyamines in Biology and Medicine (Vol 8 of The Biochemistry of Disease, Farber, E.; Pitot, H. C. Ed.), M. Dekker, New York 1981.**
- *3.* **Zappia. V.; Pegg, A. E. Progress in Polyamine Research (Vol 250 of Advances in Experimental Medicine and Biology, Back, N.; Cohen, I. R.; Kritchevsky D.; Lajtha, A.; Paoletti, R., Ed.), Plenum Press, New York 1988.**
- *4.* **Bloomfield, V. A.; Wilson, R. W. Interaction of Polyamines with Polynucleotides in ref. 2, p. 183-206.**
- *5.* **Feuerstein, B. G.; Basu, H. S.; Marton, L. J. Theoretical and Experimental Characterization of Polyamine/DNA Interactions in ref. 3, p.517-523.**
- *6.* **Tetzlaff, C.; Vilsmeier, E.; Schlag, W.-R.** *Tetrahedron,* **1990,** *46, 8117-8130.*
- *7.* **Vilsmaier, E.; Tetzlaff, C., Butz, V.; Maas, G.** *Tetrahedron,* **1991, 47, 8133-8144.**
- *8.* **Butz, V.; Vilsmaier, E.** *Tetrahedron,* **1993, 49, 6031-6044.**
- *9.* **Tetzlaff, C.; Butz, V.; Vilsmaier, E.; Wagemann, R.; Maas, G.; Ritter v. Onciul, A.; Clark, T.** *J. Chem. Sot. Perkin Trans. 2,* **1993, in press.**
- 10. Keller, E. SCHAKAL, Universität Freiburg (Germany), 1990
- **11. Kunze, U. R. Grundlagen der quantitativen Analyse, G. Thieme, Stuttgart 1980, p. 81-88.**
- 12. Wagemann, R.; Vilsmaier, E.; Fröhlich, K.; Seibel, J.; Maas, G. to be published.
- *13.* **Soloway, S.; Lipschitz, A.** *J. Org. Chem.,* **1958, 23, 613-615.**
- **14. Harmony, M. D.; Nandi, R. N.; Tietz, J. V.; Choe, J.-l.; Getty, S. J.; Staley,** *S.* **W.** *J. Am. Chem. Sot., 1983, 105, 3947-3951.*
- *15.* **Giinther, H. NMR-Spektroskopie, G. Thieme, Stuttgart 1983, 2nd edition, p 229-230.**
- **16. Lett, R. G.; Petrakis, L.; Ellis, A. F.; Jensen, R. K.** *J. Phys. Chem.,* **1970,** *74, 2816-2822.*
- *17.* **All calculations were done with the Structure Determination Package (Enraf-Nonius, Delft, The Netherlands).**
- **18. Atomic co-ordinates, bond lengths and angles, and thermal parameters have been deposited at the Cambridge Crystallographic Data Centre, University Chemical Laboratory,** Lensfield, Cambridge, CB2 1EW. The X-ray data are available on request from the Director **of the CCDC by quoting the full literature citation of this paper.**